

## Review Article

# The Atypical Lymphocyte

Michael W. Simon, MD, PhD

### Abstract

The atypical lymphocyte is a reactive lymphocyte that plays an important role in the immune response. Blood analysis is moving toward automated differentials which may miss atypical lymphocytes. The presence and number of atypical lymphocytes are useful information and in certain situations diagnostic of specific disease states. *Int Pediatr.* 2003;18(1):20-22.

**Key words:** atypical lymphocyte, automated differentials, immune response

### Introduction

With our current state of the art automated complete blood count and differential, the number or even presence of atypical lymphocytes is being under reported. We are losing important and, at times, vital information helpful in diagnosing different disease states in children. The atypical lymphocyte is a nonmalignant leukocyte seen in the peripheral blood. It is a reactive lymphocyte of lymphoid origin and produced in a variety of disorders (Table 1).<sup>1-11</sup> It appears to be a nonspecific response to stress from a variety of stimuli. A small lymphocyte becomes larger in size and capable of dividing.<sup>12</sup> It was originally described by Türk<sup>13</sup> ("Türk cell") in 1907 in the peripheral blood of patients with infectious mononucleosis and later classified in greater detail by Downey and McKinlay in 1923.<sup>14</sup> Today, it is called an atypical lymphocyte, Downey cell, or reactive lymphocyte and is a marker of illness when observed in a peripheral blood smear.

### Morphology

Atypical lymphocytes vary in morphologic detail as well as surface marker characteristics showing that

From the Department of Pediatrics, University of Kentucky, Lexington, KY (Dr Simon).

Address reprint requests to 2647 Regency Road, Lexington, KY 40503 (Dr Simon).

they comprise a heterogeneous mixture of cell types. This is the result of a polyclonal immune response to antigenic stimulation.<sup>4</sup> Morphology may differ from one case to another.<sup>4</sup> These lymphocytes are readily identifiable by their increased size and presence of active DNA synthesis.<sup>4</sup> They may look like a cross between a plasma cell and a lymphocyte and called a Plymphocyte, lymphocytoid plasma cell, or plasmacytoid lymphocyte.

Light microscopy shows that atypical lymphocytes vary in size and staining properties.<sup>2,3,5</sup> They are larger than a mature lymphocyte.<sup>2,5,15</sup> The cell may be indented at its periphery by the surrounding cells producing a scalloped appearance.<sup>5</sup> The cytoplasm is abundant and often vacuolated giving it a foamy appearance. When stained with either Wright's or Giemsa stain, the cytoplasm will vary in color being basophilic, dark blue, plasmacytic to pale gray.<sup>2,5,15</sup> It may be dark blue at the periphery of the cell. The nucleus is distinctive, often lobulated or indented. It may be oval, round, kidney-shaped, or divided and placed eccentrically.<sup>5,15</sup> It has slightly finer chromatin compared to a small lymphocyte.<sup>15</sup>

Histochemistry shows reactive lymphocytes to have increased concentrations of acid phosphatase, phosphorylase, and non-specific esterase.<sup>17,18</sup> They have a high glycogen content.<sup>18</sup> Pyroninophilia was weak and acridine orange fluorescence was absent indicating low cytoplasmic RNA content. Increased levels of formate-activating enzymes N<sup>5</sup> N<sup>10</sup> methyltetrahydrofolic dehydrogenase and dihydrofolic reductase occurred in atypical lymphocytes. Glucose-6 phosphate dehydrogenase activity is reduced.<sup>16</sup> This information is consistent with reactive lymphocytes synthesizing DNA.<sup>17</sup>

Electron microscopy has revealed a number of ultramicroscopic properties of atypical lymphocytes.<sup>4,17</sup> Endoplasmic reticulum, the Golgi apparatus and other organelles show different degrees of development. The endoplasmic reticulum is sparse. There is finely reticular nuclear chromatin, long thin frequently

**Table 1 - Causes of Atypical Lymphocytosis**

<b>Infection</b>	<b>Drug and Toxic Reactions</b>	<b>Autoimmune Disorders</b>
Epstein-Barr virus	Hydantoin drugs (Dilantin)	Rheumatoid arthritis
Cytomegalovirus	Para amino salicylic acid (PAS)	Idiopathic thrombocytopenia purpura
Toxoplasma	Phenothiazine	Systemic lupus erythematosus
Q fever	Organic arsenicals	Autoimmune hemolytic anemia
Rubella	Lead	Chronic hepatitis
Roseola	Trinitrotoluene	Agammaglobulinemia
Herpes simplex	Diaminodiphenyl-sulfone (dapsona)	
Hemorrhagic fever		<b>Malignant Disease</b>
Herpes zoster		Hodgkin's Disease
Rickettsial pox	<b>Postperfusion syndrome</b>	
Rubeola		<b>Idiopathic Disorders</b>
Mumps	<b>Immunizations</b>	Sarcoidosis
Adenovirus		Carcinomatous neuropathy
Influenza	<b>Radiation</b>	Guillain-Barre syndrome
Tuberculosis		Myasthenia gravis
Varicella	<b>Hormonal Causes</b>	Acute disseminated encephalomyelitis
Syphilis	Stress (epinephrine)	
HIV (1+2)	Addison's disease	<b>Graft Rejection</b>
Hepatitis A	Deficiencies of glucocorticoids	Renal
Hepatitis B	Panhypopituitarism	
Adenovirus	Thyrotoxicosis	
Listeria monocytogenes		
Dengue hemorrhagic fever		
Mycoplasma pneumonia		

branched mitochondria, and a well-developed Golgi apparatus.<sup>12,17</sup>

There are prominent clusters and rosettes of free ribosomes.<sup>12</sup> Some cells have many cytoplasmic RNA granules.<sup>17</sup> Small vacuoles near the edge of the cytoplasm as well as invaginations in the cell surface suggests micropinocytosis. This is seen in cells undergoing DNA synthesis. Autoradiographic studies using tritiated thymidine confirms atypical lymphocytes are actually synthesizing deoxyribonucleic acid.<sup>19-21</sup> Similarly tritiated cytidine shows reactive lymphocytes are synthesizing RNA.<sup>20</sup>

### Function

The ubiquity of the reactive lymphocyte suggests it plays an important role in the immune response. The atypical lymphocytes have been best studied from blood of patients with infectious mononucleosis. A variety of early studies using incorporation and radioautography with tritiated thymidine and rosette formation demonstrated that atypical lymphocytes are heterogeneous having both T and B cell types.<sup>4,6,22</sup> Their heterogeneity in cells suggests they are reactive in nature. These studies also confirm that they are actively proliferating.<sup>15</sup> Antigenic stimulus leads to a polyclonal proliferation lymphoid response.<sup>4,6</sup>

More recent studies suggest the reactive lymphocytes are activated T-lymphocytes produced in response to infected B-lymphocytes.<sup>23,24</sup> Atypical lymphocytes are rosette forming with sheep red blood cells confirming they are T lymphocytes that have undergone transformation.<sup>23,24,26</sup> It is possible that some of these cells identified as B-cells are actually T-cells coated with autoantibody. However, a study by Pattengale et al showed that atypical lymphocytes had sheep erythrocyte receptors as well as human T-cell specific antigens and lacked B-cell receptors, i.e., complement receptors, surface immunoglobulin, and human B-lymphocyte specific antigens.<sup>27</sup>

Reactive lymphocytes have been found to accumulate in areas of inflammation like the liver and pharynx of individuals with infectious mononucleosis and skin window preparations.<sup>28</sup> They act as normal lymphocytes in sites of local inflammation, playing a role in the immune response in a primary cellular immune or helper T-cell response. Cell-mediated immunity is important in the host defense against viral infections and control of malignant neoplasia. In infectious mononucleosis, the atypical lymphocytes are one component of a normal immune system that helps to control potentially fatal Epstein-Barr virus-induced B-cell lymphoma in man.<sup>29-31</sup> They are produced by antigenic challenge by infections, after immunizations,

rheumatoid arthritis, and systemic lupus erythematosus.<sup>19</sup>

## References

1. Tsaparas YF, Brigden ML, Mathias R, et al. Proportion Positive for Epstein-Barr Virus, Cytomegalovirus, Human Herpesvirus-6, Toxoplasma, and Human Immunodeficiency Virus Types 1 and 2 in Heterophile-Negative Patients with an Absolute Lymphocytosis or an Instrument-Generated Atypical Lymphocyte Flag. *Arch Pathol Lab Med.* 2000;124:1324-30.
2. Infectious Diseases and their Etiologic Agents. In: Mandell, Douglas, and Bennett's *Principles and Practice of Infectious Diseases*. 4th Edition. eds Mandell GL, Bennett JE, Dolin R. Churchill Livingstone, New York, 1995, pg 1370.
3. Reiss RF. Laboratory Diagnosis of Lymphoid Disorders. In: *Clinical Laboratory Medicine*. eds. Tilton RC, Balows A, Hohnadel DC, Reiss RF. Mosby Yearbook 1992, St. Louis, MO. pg 961-3.
4. Shifan TA, Mendelsohn J. The circulating "Atypical" Lymphocyte. *Hum Pathol.* 1978;9:51-61.
5. Chin T DY. Diagnosis of Infectious Mononucleosis. *South Med J.* 1976;69:654-8.
6. Giuliano VJ, Jasin HE, Ziff M. The Nature of the Atypical Lymphocyte in Infectious Mononucleosis. *Clin Immunol Immunopathol.* 1974;3:90-8.
7. *Infections - General Aspects in Hematologic Manifestations of Childhood Diseases*. Ed Lascari AD. Thieme-Stratton, Inc., New York, 1984. pg 9-10.
8. Thisyakorn U, Nimmannitya S, Ningsanon V, Soogarun S. Atypical Lymphocyte in Dengue Hemorrhagic Fever. Its Value in Diagnosis. *Southeast Asian J Trop Med Pub Hlth.* 1984;15:32-6.
9. Evans AS. Infectious Mononucleosis and Related Syndromes. *Am J Med Sci.* 1978;276:325-39.
10. Lascari AD, Bapat VR. Syndromes of Infectious Mononucleosis. *Clin Pediatr.* 1970;9:300-5.
11. Cook SD, Dowling PC. Neurological Disorders Associated with Increased DNA Synthesis in Peripheral Blood. *Arch Neurol.* 1968;19:583-90.
12. Inman DR, Cooper EH. The Relation of Ultrastructure to DNA Synthesis in Human Leukocytes. *Acta Haemat.* 1965;33:257-78.
13. Türk W. Septische Erkrankungen Bei Verkümmerng DES Granulozytensystems. *Wien Klin Wchenschr.* 1907;20:157-62.
14. Downey H, McKinlay CA. Acute Lymphadenitis Compared with Acute Lymphatic Leukemia. *Arch Intern Med.* 1923;32:82-112.
15. Peripheral Blood and Bone Marrow Assessment. In: *Clinical Laboratory Medicine* ed McClatchey KD. Williams and Wilkins, Baltimore, 1994. pg 857-8.
16. Bertino JR, Simmons BM, Donohue DM. Increased Activity of Some Folic Acid Enzyme Systems in Infectious Mononucleosis. *Blood.* 1962;19:587-92.
17. Wood TA, Frenkel EP. The Atypical Lymphocyte. *Am J Med.* 1967;42:923-36.
18. Galbraith P, Mitus WJ, Gollerkeri M, Dameshek W. The "Infectious Mononucleosis Cell" A Cytochemical Study. *Blood.* 1963;22:630-8.
19. Crowther D, Fairley GH, Sewell RL. Significance of the Changes in the Circulating Lymphoid Cells in Hodgkin's Disease. *Br Med J.* 1969;2:473-7.
20. Epstein LB, Brecher G. DNA and RNA Synthesis of Circulating Atypical Lymphocytes in Infectious Mononucleosis. *Blood.* 1965;25:197-203.
21. Horwitz DA, Stastny P, Ziff M. Circulating Deoxyribonucleic Acid-Synthesizing Mononuclear Leukocytes. I. Increased Numbers of Proliferating Mononuclear Leukocytes in Inflammatory Disease. *J Lab Clin Med.* 1970;76:391-402.
22. Greaves MF, Brown G, Rickinson AB. Epstein-Barr Virus Binding on Lymphocyte Subpopulations and the Origin of Lymphoblasts in Cultured Lymphoid Cell Lines and in the Blood of Patients with Infectious Mononucleosis. *Clin Immunol Immunopathol.* 1975;3:514-24.
23. Thomas DB. Antibodies to Membrane Antigen(s) Common to Tlymphocytes and a Subpopulation of Lymphocytes in Infectious-Mononucleosis SERA. *Lancet.* 1972;ii:399-403.
24. Sheldon PJ, Hemsted EH, Papamichail M, Holbrow EJ. Thymic Origin of Atypical Lymphoid Cells in Infectious Mononucleosis. *Lancet.* 1973;ii:1153-5.
25. Sheldon PJ, Holbrow EJ. H-Rosette Formation in T-cell-Proliferative Diseases. *Br Med J.* 1975;4:381-5.
26. Virolainen M, Andersson LC, Lalla M, Von Essen R. T-Lymphocyte Proliferation in Mononucleosis. *Clin Immunol Immunopathol.* 1973;2:114-20.
27. Pattengale PK, Smith RW, Perlin E. Atypical Lymphocytes in Acute Infectious Mononucleosis. Identification by Multiple T- and B-lymphocyte Markers. *N Engl J Med.* 1974;291:1145-8.
28. Paine TF. Atypical Lymphocytes in Throat Exudate of Patients with Infectious Mononucleosis. *N Engl J Med.* 1961;264:240.
29. Pagano JS, Huang CH, Levine P. Absence of Epstein-Barr Viral DNA in American Burkitt's Lymphoma. *N Engl J Med.* 1973;289:1395-9.
30. Dameshek W. *Speculations on the Nature of Infectious Mononucleosis. Infectious Mononucleosis*. Ed. Carter RL, Penman HG. Alden and Monbray LTD, Oxford, 1969, pg 225-40.
31. Smith RT, Bauscher JC. Epstein-Barr Virus Infection in Relation to Infectious Mononucleosis and Burkitt's Lymphoma. *Ann Rev Med.* 1972;23:39-56.